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For: **NOVEL PEPTIDES WITH INCREASED + CHARGE AND HYDROPHOBICITY BY SUBSTITUTING ONE OR MORE AMINO ACIDS OF CA-MA PEPTIDE AND PHARMACEUTICAL COMPOSITIONS CONTAINING THEREOF**

TRANSLATION OF PRIORITY DOCUMENT

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September 19, 2003

SIR:

Attached is an English language translation of the foreign priority document of the above-captioned patent application: Korean Patent Application No. 2001-57837, together with a declaration confirming the accuracy of the translation.

Respectfully submitted,

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VERIFIED STATEMENT ON TRANSLATION

I, Won-hee Lee of 8th Fl., Sung-ji Heights II, 642-16 Yeoksam-dong, Gangnam-gu, Seoul 135-080, Republic of Korea, hereby declare that I am knowledgeable in the English and Korean languages, and that to the best of my knowledge the attached document is a true and complete English translation of Korean Patent Application No. 2001-57837 filed on September 19, 2001.

Dated this 1st day of September, 2003

Won-hee Lee
Patent Attorney

Novel peptides with increased + charge and hydrophobicity by substituting one or more amino acids of CA-MA peptide and pharmaceutical compositions containing thereof

FIELD OF THE INVENTION

The present invention relates to novel peptides with increased + charge and hydrophobicity by substituting one or more amino acids of CA-MA peptide in which cecropin A (CA) and magainin 2 (MA) were conjugated and pharmaceutical compositions containing thereof. More precisely, the present invention relates to synthetic peptides prepared by substituting one or more amino acids of CA-MA peptide represented by the SEQ. ID. NO: 1 with amino acids having + charge and hydrophobicity and anti-bacterial, anti-fungal and anticancer compositions containing thereof. The synthetic peptides of the present invention have no cytotoxicity but have excellent anti-bacterial, anti-fungal and anticancer activity, leading in an effective use thereof as a safe anticancer agent and antibiotics.

BACKGROUND

Bacteria infection is one of the most common but fatal causes for human diseases. Infection has been successfully treated by antibiotics, but the abuse of

antibiotics brought another problem that bacteria now might have resistance against antibiotics. In fact, the speed which bacteria are adapting and having resistance against new antibiotics outruns that of developing new antibiotics analogues. For example, fatal *Enterococcus faecalis*, *Mycobacterium tuberculosis* and *Pseudomonas aeruginosa* are known to have raised their resistance against every possible antibiotics (Stuart B. Levy, *Scientific American*, 1998, 46-53).

Tolerance is different from resistance against antibiotics, and it was firstly found in *Pneumococcus sp.* in 1970s, which provided an important clue for disclosing the mechanism of penicillin (Tomasz, et al., *Nature*, 1970, 227, 138-140). Some bacteria species having tolerance stopped growing under the ordinary concentration of antibiotics but never died. Tolerance is caused by that the activity of autolytic enzyme of bacteria, like autolysin, is suppressed when the antibiotics inhibit cell wall synthetase. Penicillin can kill bacteria by activating endogenous hydrolytic enzyme; on the other hand, bacteria can survive by restraining the activity thereof even when being treated with antibiotics.

It is a clinical hot issue that bacteria are having tolerance against various antibiotics since

infection cannot be effectively cured with antibiotics due to the tolerance (Handwerger and Tomasz, *Rev. Infec. Dis.*, 1985, 7, 368-386). Again, once bacteria have tolerance, they can have resistance, which helps that bacteria survive under antibiotics treatment. Such bacteria can acquire new genetic elements having resistance against antibiotics, thus they can grow even under antibiotics treatment. Actually, bacteria having resistance have tolerance, too (Liu and Tomasz, *J. Infect. Dis.*, 1985, 152, 365-372). Thus, it is urgent to develop novel antibiotics, which can kill antibiotics-resistant bacteria.

There are two types of tolerance in the aspect of its mechanism. The first one is phenotypic tolerance, which occurs when the growing speed decreases in all kinds of bacteria (Tuomanen E., *Revs. Infect. Dis.*, 1986, 3, S279-S291), and the second one is genotypic tolerance acquired by mutation in a certain type of bacteria. For both cases, down regulation of autolysin activation is basically occurring. In the case of phenotypic tolerance acquired by outside stimulus, down regulation takes place temporally while down regulation occurs permanently in the case of genotypic tolerance acquired by mutation, which cause the change of hemolysis regulating routes. Autolysin deficiency is believed to cause the simplest genotypic tolerance, but

the bacteria having tolerance acquired by autolysin deficiency have not been reported yet. Such tolerance observed in clinics rather seemed to be caused by the regulation of autolysin activity (Tuomanen et al., *J. Infect. Dis.*, 1988, 158, 36-43).

In order to fight bacteria having tolerance against antibiotics, it is required to develop new antibiotics including one that is working separately from autolysin activity. In addition, it is also required to provide pharmaceutical compositions containing thereof to treat bacteria infection and inflammation effectively.

Meanwhile, bacteria can kill the neighboring bacteria by synthesizing peptides or small organic molecules, which are called bacteriocin. Such bacteriocins are classified into three groups according to their structure. The first group is lantibiotics, the second group is nonlantibiotics, and the third group is those, which are secreted by signal peptide (Cintas et al., *J. Bad.*, 1998, 180, 1988-1994). Animals including insects also produce naturally synthesized peptide antibiotics (Bevines et al., *Ann. Rev. Biochem.*, 1990, 59, 395-414), which are classified into three groups according to their structure as well. The first group is cysteine-rich β -sheet peptides, the

second group is α-helical amphiphilic peptides, and the third group is proline-rich peptides(Mayasaki et al., *Int. J. Antimicrob. Agents*, 1998, 9, 269-280). Those anti-bacterial peptides are known to play an important role in host-defense and congenital immune system (Boman, H. G., *Cell*, 1991, 65, 205; Boman, H. G., *Annu. Rev. Microbiol.*, 1995, 13, 61). The anti-bacterial peptides have many different structures depending on amino acid sequences, and the most common structure is amphiphilic α-helical structure having no cysteine, just like cecropin, an anti-bacterial peptide found in insects.

Among those peptides, the anti-bacterial activity of amphiphilic peptides has been studied and the development of antibiotics using the amphiphilic peptides has been tried. As of today, magainin 2(MA), cecropin A (CA) and melittin (ME) have been reported as amphiphilic peptides.

Amphiphilic peptides of cecropin group were first found in a fruit fly and later in a silkworm pupa and in a pig intestine, too. While cecropin A was reported to have high anti-bacterial activity but low anti-fungal and anticancer activity (Boman, H. G. and Hultmark, D., *Annu. Rev. Microbiol.*, 1987, 41, 103), magainin 2 was known not to have cytotoxicity but to have appreciable anti-bacterial, anti-fungal, anticancer and anti-protozoa activity (Zasloff, M.,

Proc. Natl. Acad. Sci. USA, 1987, 84, 5449). It has been further reported that new synthetic peptides having excellent anti-bacterial, anti-fungal and anticancer activity could be prepared by constructing conjugation peptide recombinated some parts of the sequences of the above two peptides (Chan, H. C., et al., *FEBS Lett.*, 1989, 259, 103; Wade, D., et al., *Int. J. Pept. Prot. Res.*, 1992, 40, 429).

The present inventors have designed and synthesized novel peptides having amino acid sequences with + charge and hydrophobicity at amino terminal, taking amphiphilic peptide conjugated cecropin A and magainin 2 as a template. And the present invention has been accomplished by confirming that the synthetic peptides of the present invention could be effectively used as anticancer agents and antibiotics owing to their anti-bacterial, anti-fungal and anticancer activity.

SUMMARY OF THE INVENTION

It is an object of this invention to provide novel peptides and their derivatives with increased + charge and hydrophobicity by substituting one or more amino acids of cecropin A and magainin 2 conjugated CA-MA peptide represented by the SEQ. ID. NO: 1 and with

excellent anti-bacterial, anti-fungal and anticancer activity without cytotoxicity.

It is another object of this invention to provide pharmaceutical compositions for anti-bacterial, anti-fungal and anticancer agent containing the above synthetic peptides.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 is photographs showing the number of colonies on LB agar plate, in which *Bacillus subtilis* was treated with synthetic peptide of the present invention,

A: Positive control,

B: CA-MA peptide,

C: Synthetic peptide represented by the SEQ. ID. NO: 2

FIG. 2 is photographs showing the number of colonies on NB+0.5% NaCl agar plate, in which *Pseudomonas aeruginosa* was treated with synthetic peptide of the present invention,

A: Positive control,

B: CA-MA peptide,

C: Synthetic peptide represented by the SEQ. ID. NO: 2

FIG. 3 is SEM (scanning electron microscopy) microphotographs showing the result of treating synthetic peptide of the present invention to *Bacillus subtilis*,

A: Positive control,

B: CA-MA peptide,

C: Synthetic peptide represented by the SEQ. ID. NO: 2

FIG. 4 is SEM microphotographs showing the result of treating synthetic peptide of the present invention to *Pseudomonas aeruginosa*,

A: Positive control,

B: CA-MA peptide,

C: Synthetic peptide represented by the SEQ. ID. NO: 2

FIG. 5 is graphs showing the dynamic condition of lipid membrane after treating synthetic peptide of the present invention to *Bacillus subtilis* and *Pseudomonas aeruginosa*,

A: Dynamic condition of lipid membrane of *Bacillus subtilis*,

B: Dynamic condition of lipid membrane of *Pseudomonas aeruginosa*,

● : CA-MA peptide,

□ : Synthetic peptide represented by the SEQ. ID.

NO: 2

FIG. 6 is graphs showing the anticancer activity of the synthetic peptide of the present invention against various cancer cell lines.

A: Anticancer activity against Calu-6 cell line,

B: Anticancer activity against Jurkat cell line,

C: Anticancer activity against SNU 601 cell line,

● : CA-MA peptide,

0 : Synthetic peptide represented by the SEQ. ID. NO: 2

DETAILED DESCRIPTION OF PREFERRED EMBODIMENTS

To accomplish those objects, the present invention provides novel peptides and their derivatives with increased + charge and hydrophobicity by substituting one or more amino acids of cecropin A and magainin 2 conjugated CA-MA peptide represented by the SEQ. ID. NO:1.

The present invention also provides anti-bacterial, anti-fungal and anticancer pharmaceutical compositions containing the above peptides and their derivatives.

Hereinafter, the present invention is described in detail.

In one aspect, the present invention provides

novel peptides and their derivatives with increased + charge and hydrophobicity by substituting one or more amino acids of cecropin A and magainin 2 conjugated CA-MA peptide represented by the SEQ. ID. NO: 1.

Peptides and their derivatives of the present invention were synthesized to have increased + charge and hydrophobicity by substituting a few amino acids including hinge region of CA-MA peptide which was prepared by conjugating 1-8 amino acid region of amphiphilic helical CA and 1-12 amino acid region of MA, and represented by the SEQ. ID. NO: 1, with other amino acids.

In order to prepare synthetic peptides of the present invention, the present inventors used Merrifield's liquid solid phase method in which Fmoc (9-fluorenylmethoxycarbonyl) was used as a protecting group (Merrifield, R. B., *J. Am. Chem. Soc.*, 1963, 85, 2149). Every synthetic peptide with increased + charge and hydrophobicity by substituting one or more amino acids including hinge region of CA-MA peptide represented by the SEQ. ID. NO: 1 could be peptide of the present invention. Especially, peptides and their derivatives prepared by substituting glysine-isoleucine-glycine residing at hinge region of CA-MA peptide represented by the SEQ. ID. NO: 1 with proline each, substituting 4^{th} leucine, 8^{th} isoleucine, 14^{th} leucine, 15^{th} histidine with lysine each, and

substituting 5th phenylalanine, 6th lysine, 12th lysine, 13th phenylalnine, 16th serine, 17th alanine, 20th phenylalanine with leucine were preferred.

The peptide synthesized as above was isolated and purified, after which the purity thereof was confirmed. As a result, the purity of the peptide was over 95%, and the molecular weight obtained by MALDI (Matrix-Assisted Laser Desorption Ionization) mass spectrometry (Hill, et al., *Rapid Commun. Mass Spectrometry*, 1991, 5, 395) was the same as the molecular weight obtained by calculation of amino acids. Therefore, it was confirmed that the peptide having correct amino acid sequence represented by the SEQ. ID. NO: 2 was synthesized.

The present invention also provides anti-bacterial, anti-fungal and anticancer pharmaceutical compositions containing the above peptides and their derivatives.

To confirm if the peptides and their derivatives of the present invention can be used for anti-bacterial, anti-fungal and anticancer agents, the present inventors have measured the anti-bacterial activity of the synthetic peptides by measuring minimal inhibitory concentration (referred as "MIC" hereinafter).

Synthetic peptide of the present invention represented by the SEQ. ID. NO: 2 was used to measure MIC value to each strain. As a result, synthetic

peptide of the present invention was confirmed to have more than 4-fold anti-bacterial activity (varied a little depending on strains), comparing to the comparative group using CA-MA conjugation peptide (see Table 1).

Also, anti-bacterial activity of the peptide of the present invention against *Bascilus subtilis* and *Pseudomonas aeruginosa* was measured on LB agar plate. As a result, synthetic peptide of the present invention represented by the SEQ. ID. NO: 2 was confirmed to have remarkable anti-bacterial activity comparing to the CA-MA conjugation peptide (see Fig.1 and Fig. 2).

In addition, observing the anti-bacterial activity of the present invention against *Bascilus subtilis* and *Pseudomonas aeruginosa* with scanning electron microscopy also supported the same result as above (see Fig. 3 and Fig. 4).

Again, synthetic peptide of the present invention represented by the SEQ. ID. NO: 2 was confirmed to have remarkable anti-bacterial activity comparing to the comparative group using CA-MA conjugation peptide, which was resulted from observing the dynamic condition of lipid membrane after *Bascilus subtilis* and *Pseudomonas aeruginosa* were treated with the synthetic peptide (see Fig. 5).

In order to measure the anti-fungal activity of

synthetic peptide of the present invention, the MIC values to *Candida albicans* and *Trichosporon beigelii* were measured by MTT assay method. As a result, the synthetic peptide of the present invention represented by the SEQ. ID. NO: 2 showed more than 2-fold anti-fungal activity comparing to the comparative group using CA-MA peptide (see Table 2).

In order to see if the synthetic peptide of the present invention have anticancer activity, human lung cancer cell line Calu-6, human stomach cell line SNU 601 and T-cell lymphoma cell line were treated with the peptide. As a result, the synthetic peptide of the present invention represented by the SEQ. ID. NO: 2 was confirmed to have higher anticancer activity than the comparative group using CA-MA peptide (see Fig. 6).

Further, the present inventors measured the hemolysis capacity of the synthetic peptide of the present invention in order to see if it has cytotoxicity. As a result, along with CA-MA peptide, the synthetic peptide of the present invention represented by the SEQ. ID. NO: 2 had no cytotoxicity. Meanwhile, melittin, bee venom, used as a positive control showed high cytotoxicity (see Table 3).

Considering all those results together, the

synthetic peptide of the present invention represented by the SEQ. ID. NO: 2 was confirmed to have excellent anti-bacterial, anti-fungal and anticancer activity without cytotoxicity, so that the peptide can be effectively used as a safe anti-bacterial, anti-fungal and anticancer treatment agent.

Peptides and their derivatives of the present invention can be administered orally or parenterally. The compounds of the present invention can be prepared for oral or parenteral administration by mixing with generally-used fillers, extenders, binders, wetting agents, disintegrating agents, diluents such as surfactant, or excipients. Formulations for parenteral administration are sterilized aqueous solutions, water-insoluble excipients, suspensions, emulsions, and suppositories. Water-insoluble excipients and suspension can contain propylene glycol, polyethylene glycol, vegetable fats such as olive oil and injectable ester such as ethyl oleit. Suppositories can contain, in addition to the active compound, witepsol, macrogol, tween 61, cacao oil, laurin oil and glycerogelatin.

Also, peptides and their derivatives of the present invention can be used mixing with pharmaceutically acceptable carriers such as physiological saline or organic solvent, and with carbohydrate such as glucose, sucrose or dextran,

antioxidants such as ascorbic acid or glutathione, chelating agents, small molecular protein or other stabilizers for increasing stability and desorption.

In general, it has proved advantageous both in human and in veterinary medicine to administer the active compound or compounds according to the present invention in total amounts of about 0.1 to about 2 mg/kg, preferably 0.5 to 1 mg/kg of body weight, 1-3 times every 24 hours, if appropriate in the form of several individual doses, to achieve the desired results. However, it may be necessary to deviate from the dosages mentioned, and in particular to do so as a function of the nature and body weight of the object to be treated, the nature and severity of the disease, the nature of the formulation and of the administration of the medicament and the period or interval within which administration takes place. Thus in some cases it can suffice to manage with less than the abovementioned amount of active compound, while in other cases the abovementioned amount of active compound must be exceeded. The particular optimum dosage and mode of administration required for the active compounds can be determined by any expert on the basis of his expert knowledge.

EXAMPLES

Practical and presently preferred embodiments of the present invention are illustrative as shown in the following Examples.

However, it will be appreciated that those skilled in the art, on consideration of this disclosure, may make modifications and improvements within the spirit and scope of the present invention.

Example 1: Synthesis of oligonucleotide represented by the SEQ. ID. NO: 1

In order to synthesize the peptide of the present invention represented by the SEQ. ID. NO: 2, we, the present inventors used Merrifield's liquid solid phase method in which Fmoc (9-fluorenylmethoxycarbonyl) was used as a protecting group (Merrifield, R. B., *J. Am. Chem. Soc.*, 1963, 85, 2149). For the peptide having $-NH_2$ type carboxyl terminal, rink amide MBHA-resin was used as a starting material. And, Fmoc-amino acid-Wang resin (SynPep Corporation) was used for the peptide having -OH type carboxyl terminal. The extension of peptide chain by Fmoc-amino acid coupling was performed by N-hydroxybenzo triazole (HOBt)-dicyclohexycarbodiimide (DCC) method. Particularly, Fmoc-

amino acid of amino terminal of each peptide was coupled, and the Fmoc group was removed by using 20% piperidine/NMP (N-methyl pyrolidone) solution. After washing with NMP and DCM (dichoromethane), the peptide was dried with nitrogen gas. TAF (trifluoroacetic acid)-phenol-thioanisole-H_2O-triisopropylsilane (85: 5: 5: 2.5: 2.5 vol/vol) solution was added thereto. In order to remove protecting group and to separate peptide from resin, the peptide was reacted for 2-3 hours, and it was precipitated by using diethylether. The crude peptide was purified by using reverse phase(RP)-HPLC column(Delta Pak, C_{18} 300 Å, 15, 19.0 ㎜ × 30 ㎝, Waters) in acetonitrile gradient containing 0.05% TFA. Synthesized peptide was hydrolyzed with 6 N-HCl at 110℃, and the residues were vacuum concentrated. And then, its amino acid composition was analyzed with amino acid analyzer (Hitachi 8500 A) after dissolving in 0.02 N-HCl. As a result, the purity of the peptide was over 95%, and the molecular weight obtained by MALDI mass spectrometry (Hill, et al., *Rapid Commun. Mass Spectrometry*, 1991, 5, 395) was the same as the molecular weight obtained by calculation of amino acids. Therefore, it was confirmed that the peptide having correct amino acid sequence was synthesized.

Experimental example 1: Anti-bacterial activity of the peptides

<1-1> Measurement of MIC

In order to measure the anti-bacterial activity of the peptide synthesized in Example 1, minimum inhibitory concentration (MIC) of the peptide was measured.

The present inventors used *Bacilus subtilis* (KCTC 1918) and *Stapilococus epidermidis* (KCTC 1917) as Gram-positive bacteria, and *Pseudomonas aeruginosa* (KCTC 1637) and *Salmonella typhimurium* (KCTC 1926) as Gram-negative bacteria for this experiment. All the bacteria used in this experiment were given by Korea Research Institute of Bioscience and Biotechnology (KRIBB). Each bacteria strain was cultured in LB medium(1% bacto-trypton, 0.5% bacto yeast extract, 1% sodium chloride) to the mid-log phase, and diluted with 1% bacto-peptone medium at the concentration of 1×10^4 cells/100 $\mu\ell$. The diluted bacteria were loaded into micro-titrate plate. Antibiotic peptide synthesized in Example 1 and CA-MA peptide(as a comparative group) were half-fold diluted consecutively from 25 μ M/well, and added into the plate for 6-hour culture at 37℃. Finally, MIC of each strain was determined by observing OD_{620} with micro-titrate plate reader. The results were

described in Table 1.

<Table 1>

Anti-bacterial activity of peptides against Gram-positive and Gram-negative bacteria

Peptide	MIC(μ M)			
	Gram-positive		Gram-negative	
	B. subtilis	*S. epidermidis*	*P. aeruginosa*	*S. typhimurium*
CA-MA	3.12	3.12	1.56	0.19
Synthetic peptide(SEQ .ID.NO:2)	0.78	1.56	0.78	0.097

As a result, it was confirmed that the antibiotic peptide of the present invention represented by the SEQ. ID. NO: 2 had around 4 times higher antibiotic activity than that of CA-MA peptide.

<1-2> Visualization of anti-bacterial activity

In order to visualize the antibiotic activity of the synthetic peptide of the present invention on the plate, *Pseudomonas aeruginosa* and *Bacilus subtilis* were inoculated in LB medium (1% bacto trypton, 0.5% yeast extract, 1% sodium chloride), and cultured to mid-log phase. Particularly, 4×10^5 *P. aeruginosa* cells were loaded into the medium, and 4 μ M of synthetic peptide was added thereto. 4×10^5 *B. subtilis* cells were also loaded into the medium, and 1 μ M of synthetic peptide

was added thereto. After culturing for 2 hours at 37℃, the culture fluid was smeared on LB plate to visualize the cells. At this time, CA-MA peptide was used as a comparative group.

As a result, lots of colonies were found in positive control group (Fig. 1A), colonies were found to be a little grown in a group where CA-MA peptide was added (Fig. 1B) and no colony was found in a group where synthetic peptide of the present invention was added (Fig. 1C), meaning the peptide of the present invention could completely inhibit the growth of bacteria.

From the above results, it was confirmed that the synthetic peptide of the present invention represented by the SEQ. ID. NO: 2 had superior antibiotic activity to that of CA-MA peptide.

<1-3> anti-bacterial activity observation with SEM

Anti-bacterial activity of the synthetic peptide of the present invention was observed with SEM (scanning electron microscopy). *Bacillus subtilis* (Gram-positive) and *Pseudomonas aeruginosa* (Gram-negative) cells were cultured in LB medium (1% bacto trypton, 0.5% bacto yeast extract, 1% sodium chloride) to mid-log phase, and the cells were diluted with 10 mM of Na-phosphate buffer containing 100 mM of NaCl) at

the concentration of 10^8 cells/mℓ. Synthetic peptide of the present invention and CA-MA peptide (as a comparative group) were added into the diluted cell culture medium (final conc. 0.78 μM in *B. subtilis* culture, 1.56 μM in *P. aeruginosa* culture), followed by further culturing for 30 minutes at 37℃. 0.2 M Na-phosphate buffer containing 5% glutaraldehyde was added into the medium, and the cells were fixed for 2 hours at 4℃. The cells were filtered with isopore filters (0.2 ㎛ pore size, Millipore, Bedford, MA, USA), and washed with 0.1 M Na-cacodylate buffer (pH 7.4). The filters were treated with 1% osmium tertoxide and dehydrated. After freeze-drying and gold coating, the filters were observed with SEM (HITACHI S-2400, Japan).

As a result, when *B. subtilis* and *S. aeruginosa* were treated with the synthetic peptide of the present invention represented by the SEQ. ID. NO: 2, much more destroyed cells were observed than when in control and when the cells were treated with CA-MA peptide (Fig. 3 and Fig. 4).

<1-4> Measurement of membrane dynamic condition

The present inventors performed the below experiment in order to investigate the dynamic condition of lipid membrane of bacteria cells treated with synthetic peptide of the present invention.

Particularly, *B. subtilis* (Gram-positive) and *P. aeruginosa* (Gram-negative) were cultured to mid-log phase in LB medium (1% bacto trypton, 0.5% bacto yeast extract, 1% sodium chloride). And the antibiotic peptide of the present invention and CA-MA peptide(as a comparative group) were treated(6.25 μM ~ 0.097 μM, half-fold diluted) thereto. Each strains were further cultured for 2 hours at 37℃. After fixing with 0.25% formaldehyde for 1 hour at room temperature, cultured cells were washed with PBS (pH 7.4), and then frozen in liquid nitrogen. For the fluorescent labeling, PBS (pH 7.4) was added until OD_{450} reached to 0.25, and DPH (1,6-diphenyl-1,3,5-hexatriene) dissolved in tetrahydrofuran was added (final conc. 10^{-4} M), followed by further culturing for 45 minutes at 37℃. Steady-state fluorescence anisotropy was determined by measuring the strength of fluorescence with spectrofluorometer (HITACHI F-3010, Tokyo, Japan) at 330 nm and 450 nm.

As a result, when *Bacillus subtilis* and *Pseudomonas aeruginosa* were treated with the synthetic peptide of the present invention represented by the SEQ. ID. NO: 2, DPH-labeled fluorescent materials were intercalated 15-20% lower position of membrane comparing to when the cells were treated with CA-MA peptide (Fig. 5).

<Experimental Example 2> Anti-fungal activity of synthetic peptide

<2-1> MTT assay

In order to measure the anti-fungal activity of the synthetic peptide of the present invention, the present inventors performed MTT assay with *Candida albicans* (TIMM 1768) and *Tricosphoron beigelii* (KCTC 7707). Particularly, PDB medium(20% potato infusion frum, 2% bacto dextrose) containing various fungi was loaded into the wells(100 ㎕/well) of 96-well plate. Antibiotic peptides of the present invention and CA-MA peptide (as a comparative group) were half-fold diluted consecutively, and added into the plate for further culturing. 10 ㎕ of MTT solution(3-[4,5-dimethyl-2-thiazolyl-2-thiazolyl]-2,5-diphenyl-2H-tetrazolium bromide, 5 ㎎/㎖) was added into each well followed by further culturing for 5-6 hours. Formazan produced by mitochondria enzymes of living cells was dissolved in 100 ㎕ of 0.04 N HCl-isopropanol. Finally, OD_{570} was measured by using ELISA reader to determine the degree of MIC. The result was described in Table 2.

<Table 2>

Anti-fungal activity of peptides

Peptide	MIC(μ M)	
	C. albicans	*T. beigelii*
CA-MA	12.5	6.25

Synthetic peptide	6.25	3.25

As a result, it was confirmed that the anti-fungal activity of the synthetic peptide of the present invention represented by the SEQ. ID. NO: 2 was about 2 times higher than that of CA-MA peptide.

<Experimental Example 3> Anticancer activity of antibiotic peptide

In order to measure the anticancer activity of the synthetic peptide of the present invention, the present inventors performed MTT assay with Calu-6 (a human lung cancer cell line), SNU 601(a human stomach cancer cell line) and Jurkat (a T-cell lymphoma cell line) cells. Firstly, 90 $\mu\ell$ of each cell line(2×10^5 cells/$m\ell$) was loaded into each well of 96-well plate. At this time, only medium contained wells were used as a control. After shaking well, the cells were cultured in CO_2 incubator for 3 days. Formazan produced by mitochondria enzymes of living cells was dissolved in 100 $\mu\ell$ of 0.04 N HCl-isopropanol, and finally, OD_{540} was measured by using ELISA reader. The anti-cancer activity of antibiotic peptide of the present invention was represented by a percentage (OD of synthetic peptide treated well/OD of control$\times$ 100).

As shown in Fig. 6, it was confirmed that the anti-cancer activity of the synthetic peptide of the

present invention was higher than that of CA-MA peptide in all cell lines. To the concentration of 1 μ M, synthetic peptide of the present invention did not showed anti-cancer activity. However, as concentration increases, the rapidly growing anticancer activity was detected. For example, strong anticancer activity which made complete restrain of cancer cell growth was observed with over 10 μ M concentration.

<Experimental example 4> Cytotoxicity of synthetic peptide

In order to confirm if the synthetic peptide of the present invention showed cytotoxicity, hemolysis capacity of the synthetic peptide was investigated.

Human red blood corpuscles were diluted with PBS (pH 7.0) to the concentration of 8%, and loaded into each wells of 96-well plate. Synthetic peptide of the present invention was half-fold diluted consecutively from 12.5 μ M/well, followed by reacting with the red blood corpuscles for 1 hour at 37℃. After centrifugation, OD_{414} was measured to determine the amount of hemoglobin in the supernatant. At this time, CA-MA peptide was used as a comparative group and melittin was used as a positive control. In order to investigate the level of hemolysis, 1% triton X-100 was added, and then OD was measured. Hemolysis capacity of triton X-100 was regarded as 100%, with which hemolysis

capacity of the synthetic peptide was compared and calculated according to the below <Mathematical Formula 1>.

<Mathematical Formula 1>

$$\% \text{ hemolysis} = (OD\ A - OD\ B / OD\ C - OD\ B) \times 100$$

In the above <Mathematical Formula 1>,

OD A = OD_{414} of peptide solution,

OD B = OD_{414} of PBS,

OB C = OB_{414} of 1% triton X-100.

The results were described in Table 3.

<Table 3>

Cytotoxicity of peptides

Peptide	% hemolysis(μ M)							
	12.5	6.25	3.125	1.56	0.78	0.39	0.195	0.097
CA-MA	0	0	0	0	0	0	0	0
Synthetic Peptide(SEQ.ID.NO:2)	0	0	0	0	0	0	0	0
Melittin	100	100	95	93	31	0	0	0

As a result, while the bee venom, melittin, showed high cytotoxicity, CA-MA peptide and synthetic peptide of the present invention represented by the SEQ. ID. NO: 2 did not show any cytotoxicity.

<Experimental Example 5> Acute toxicity test in rat via non-oral administration

The following experiments were performed to see if the synthetic peptide of the present invention has acute toxicity in rat.

6-week old SPF SD line rats were used in the tests for acute toxicity. Synthetic peptide of the present invention represented by the SEQ. ID. NO: 2 was suspended in 0.5% methyl cellulose solution and intravenous injected once to 2 rats per group at the dosage of 1 g/kg/15 mℓ. Death, clinical symptoms, and weight change in rats were observed, hematological tests and biochemical tests of blood were performed, and any abnormal signs in the gastrointestinal organs of chest and abdomen were checked with eyes during autopsy. The results showed that the synthetic peptide of the present invention did not cause any specific clinical symptoms, weight change, or death in rats. No change was observed in hematological tests, biochemical tests of blood, and autopsy. Therefore, the synthetic peptide used in this experiment are evaluated to be safe substances since they do not cause any toxic change in rats up to the level of 10 mg/kg in rats.

INDUSTRIAL APPLICABILITY

As shown above, the synthetic peptides and their derivatives of the present invention represented by the SEQ. ID. NO: 2 have no cytotoxicity but have excellent anti-bacterial, anti-fungal and anticancer activity, leading in an effective use thereof as a safe anticancer agent and antibiotics.

Those skilled in the art will appreciate that the conceptions and specific embodiments disclosed in the foregoing description may be readily utilized as a basis for modifying or designing other embodiments for carrying out the same purposes of the present invention. Those skilled in the art will also appreciate that such equivalent embodiments do not depart from the spirit and scope of the invention as set forth in the appended claims.

What is claimed is

1. Peptides or their derivatives with increased + charge and hydrophobicity by substituting one or more amino acids of CA-MA peptide represented by the SEQ. ID. NO: 1.

2. Peptides or their derivatives as set forth in claim 1, wherein the peptide is represented by the SEQ. ID. NO: 2.

3. Anti-bacterial pharmaceutical compositions containing the peptides or their derivatives of claim 1 or 2.

4. Anti-fungal pharmaceutical compositions containing the peptides or their derivatives of claim 1 or 2.

5. Anticancer pharmaceutical compositions containing the peptides or their derivatives of claim 1 or 2.

6. The pharmaceutical compositions as set forth in claim 5, wherein the cancer is lung cancer.

7. The pharmaceutical compositions as set forth in claim 5, wherein the cancer is stomach cancer.

8. The pharmaceutical compositions as set forth in claim 5, wherein the cancer is lymphoma.

ABSTRACT OF THE DISCLOSURE

The present invention relates to novel peptides with increased + charge and hydrophobicity by substituting one or more amino acids of CA-MA peptide in which cecropin A (CA) and magainin 2(MA) were conjugated and pharmaceutical compositions containing thereof. More precisely, the present invention relates to synthetic peptides prepared by substituting one or more amino acids of CA-MA peptide represented by the SEQ. ID. NO: 1 with amino acids having + charge and hydrophobicity and anti-bacterial, anti-fungal and anticancer compositions containing thereof. The synthetic peptides of the present invention have no cytotoxicity but have excellent anti-bacterial, anti-fungal and anticancer activity, leading in an effective use thereof as a safe anticancer agent and antibiotics.

Figures

FIG. 1

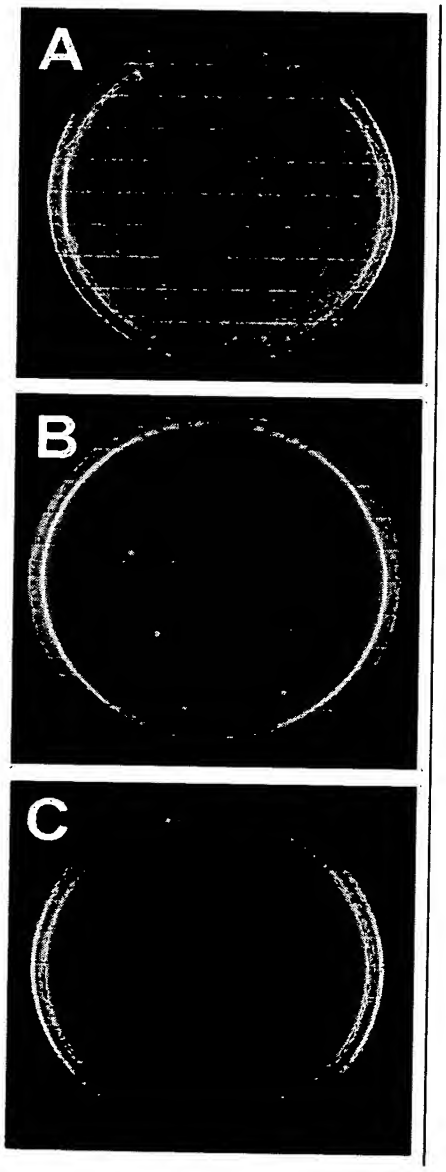


FIG.2

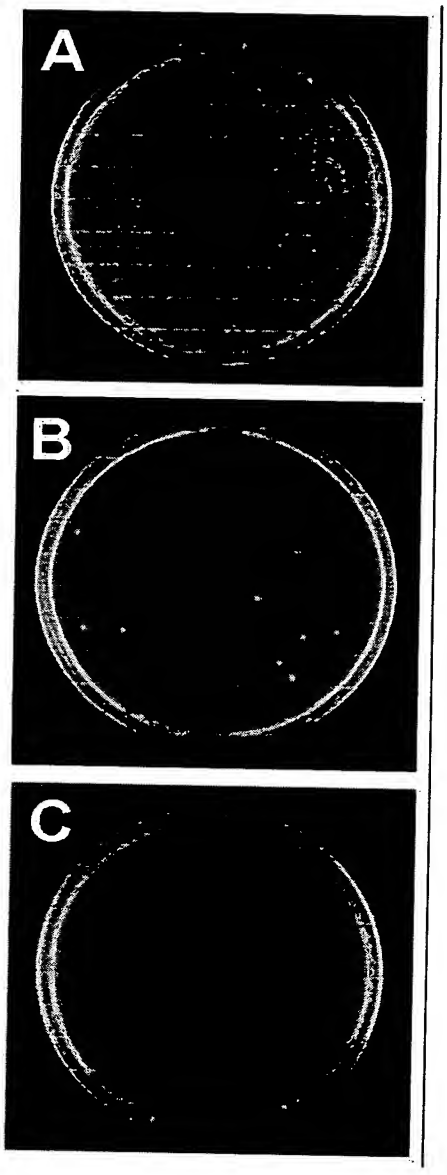


FIG. 3

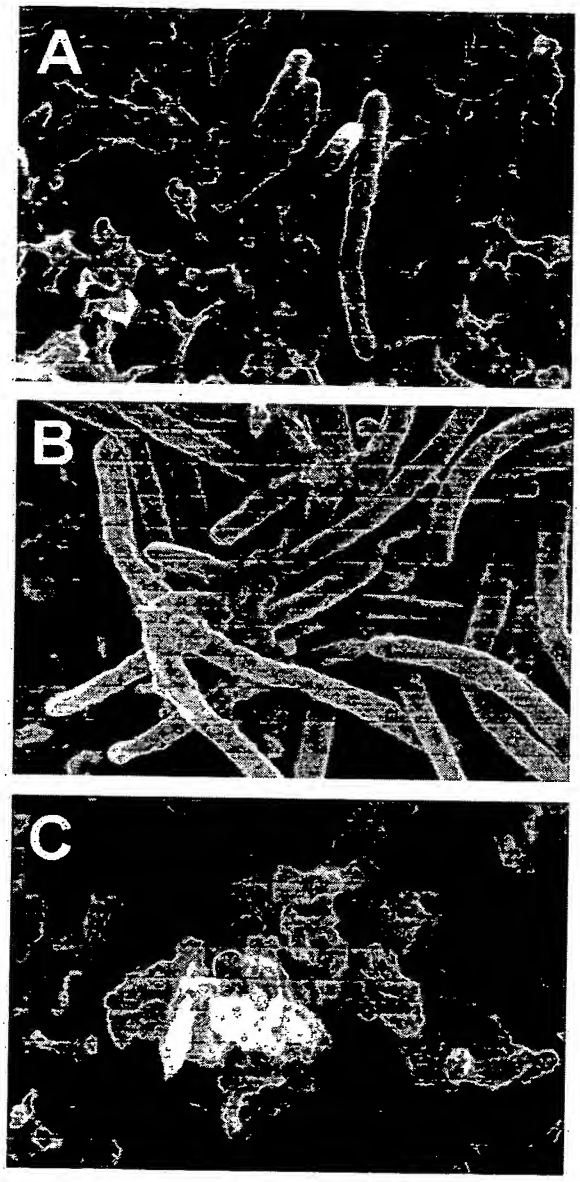


FIG. 4

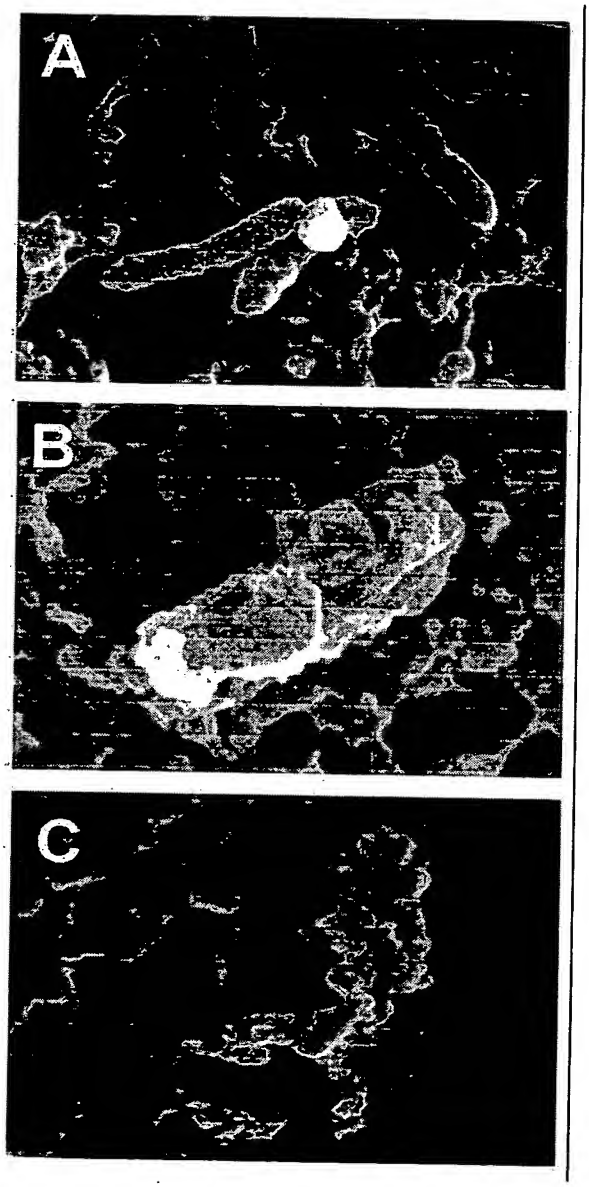
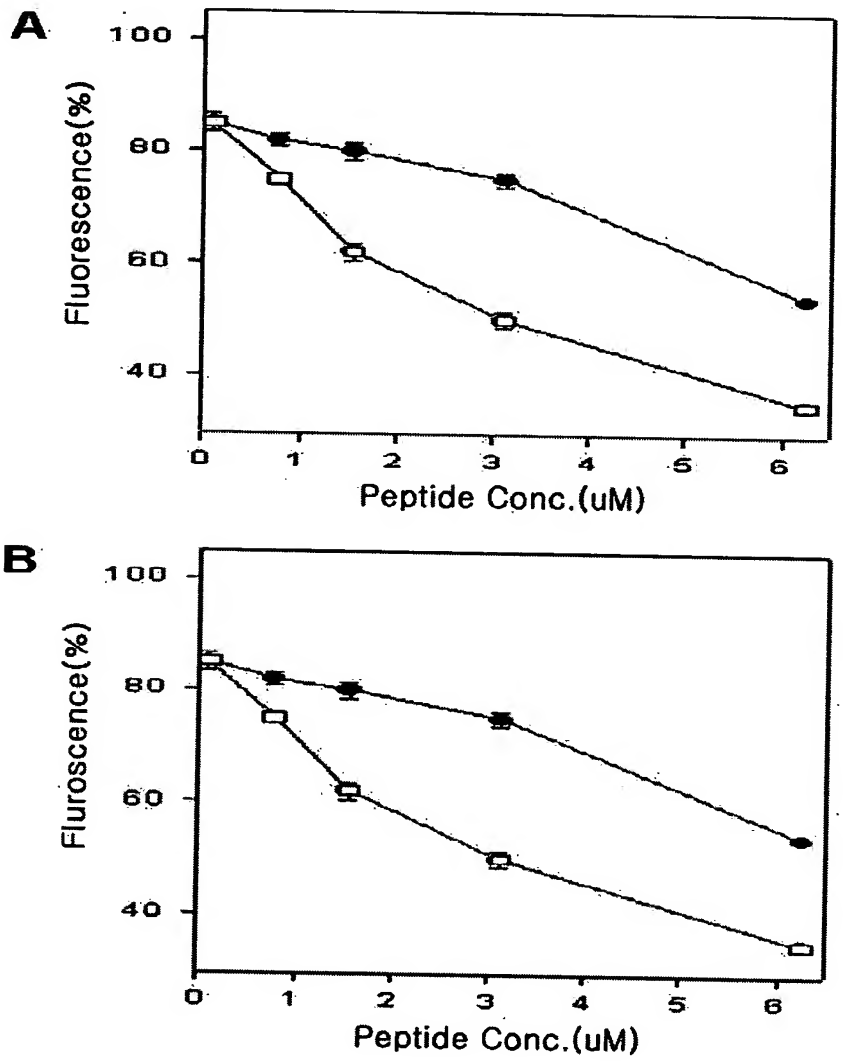


FIG. 5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60
40
Fluorescence(%)
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1
2
3
4
5
6
Peptide Conc.(uM)
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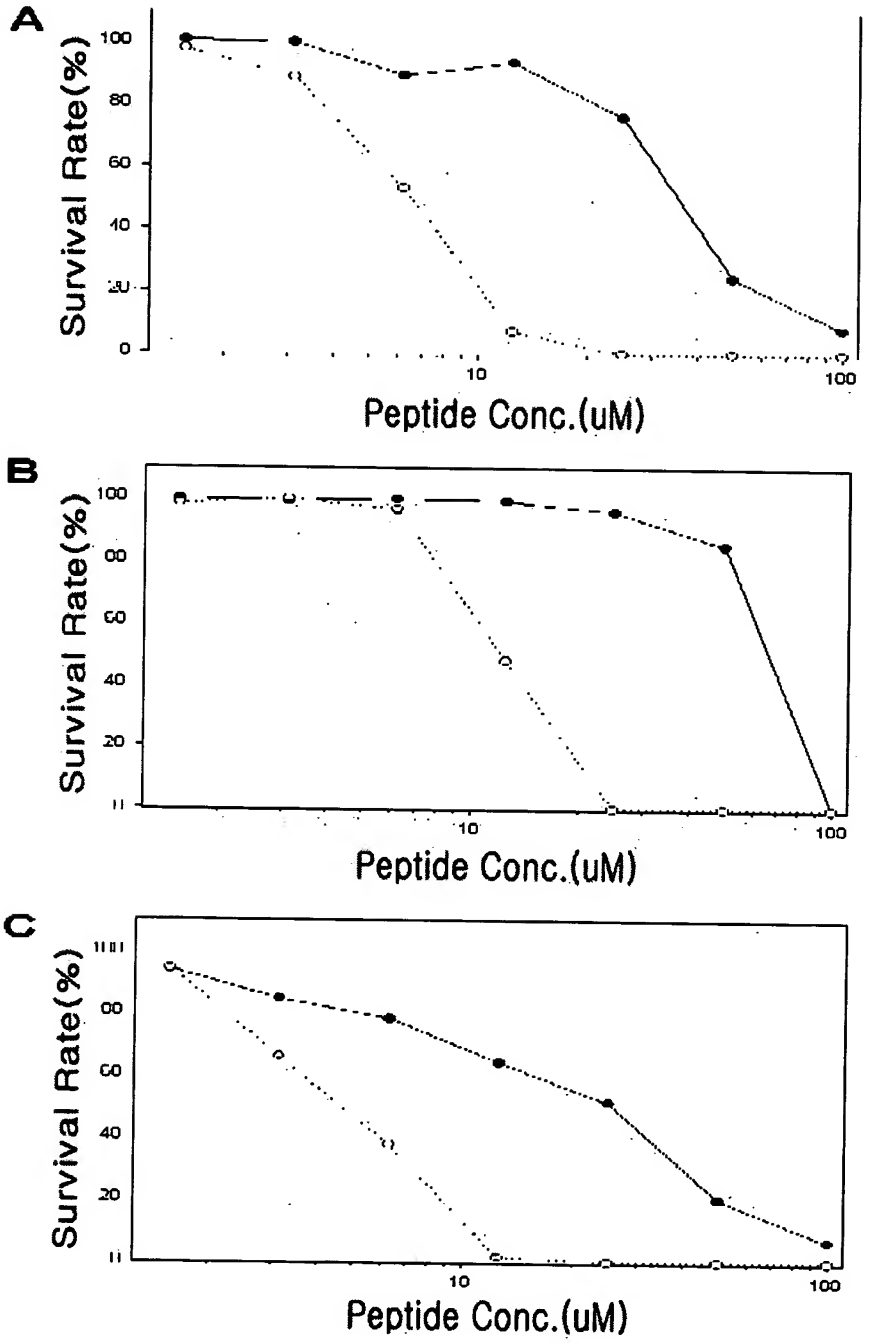
100
80
60
40
Fluroscence(%)
0
1
2
3
4
5
6
Peptide Conc.(uM)

FIG. 6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60
40
20
0
10
100
Peptide Conc.(uM)
B
Survival Rate(%)
100
80
60
40
20
10
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Survival Rate(%)
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SEQUENCE LISTING

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<120> Novel peptides with increased + charge and hydrophobicity by substituting one or more amino acids of CA-MA peptide and pharmaceutical compositions containing thereof

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Ala Lys Lys Phe
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<213> Artificial Sequence

<220>

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Leu Lys Lys Leu
20